



ESTD : 1880

**St Aloysius College (Autonomous)
Mangaluru**

**Re-accredited by NAAC “A++” Grade
Course structure and syllabus of**

**B.Sc.
BIOTECHNOLOGY
Under NEP Regulations, 2021**

ಸಂತ ಅಲೋಶಿಯಸ್ ಕಾಲೇಜು (ಸ್ವಾಯತ್ತ)

ಮಂಗಳೂರು- 575 003, ಕರ್ನಾಟಕ

www.stalloysius.edu.in



ST ALOYSIUS COLLEGE (AUTONOMOUS)

P.B. NO. 720, MANGALURU - 575 003, KARNATAKA, INDIA

Phone: +91- 0824-4117701, 4117702, 4117703, 4117704

Email: principal@stalloysius.edu.in

alloysius.principal@gmail.com

Re-accredited by NAAC with 'A++' Grade with CGPA 3.67/4 (Cycle 4)

Recognised as Centre for Research Capacity Building under UGC-STRIDE Scheme

Recognised under DBT - BUILDER Scheme, Government of India

College with "STAR STATUS" Conferred by DBT, Government of India

Recognised by UGC as "College with Potential for Excellence"

Date: 21-

02-2022

NOTIFICATION

Sub: Syllabus of **B.Sc. BIOTECHNOLOGY** under NEP Regulations, 2021.
(As per Mangalore University guidelines)

- Ref: 1. Decision of the Academic Council meeting held on 18-12-2021 vide
Agenda No: 6
2. Decision of the Academic Council meeting held on 09-07-2022 vide
Agenda No: 14
3. Decision of the Academic Council meeting held on 25-02-2023 vide
Agenda No: 3
4. Office Notification dated 21-02-2022
5. Office Notification dated 17-08-2022
6. Office Notification dated 26-09-2023

Pursuant to the above, the Syllabus of **B.Sc. BIOTECHNOLOGY** under NEP Regulations, 2021 which was approved by the Academic Council at its meeting held on 18-12-2021, 09-07-2022 & 02-09-2023 is hereby notified for implementation with effect from the academic year **2021-22**.

**PRINCIPAL
REGISTRAR**

To:

1. The Chairman/Dean/HOD.
2. The Registrar Office
3. Library

The BOS in Biotechnology (UG) held on 18th November 2021 in Conference Room, Admin Block

Following Members were present for the meeting

Chairman : Mr. Sreejesh P C

Member : Mrs. Renita Maria D'Souza

Subject experts : Dr. Raghavendra Rao, Associate Professor, Alvas College, Moodbidri

: Dr Shayama Prasad, Associate Professor, NITTE, Karkala

Vice Chancellor's nominee: Dr Tharavathy N C, Dept. of Bioscience,

Mangalore University

Representative from Industry : Mr Pavan Pai ,

Shri Laxmi Narasimha Rice & Oil Mills company,

Bhatkal, Mangalore.

Meritorious Alumnus : Dr. Lyned Lasrado

Special Invitees : Rev Dr Leo D'Souza

Dr Shreelaitha Suvarna

Dr. Shashi Kiran Nivas

Student Representative : Miss Anindita Jana, Reg No.1929 III BSc BtCZ

An online meeting (Google meet) of the Board of Study in Biotechnology (UG) was held on 21st June 2022 at 10.00 a.m., to discuss the following agenda:

Sl. No	Name	Role
1.	Mr. Sreejesh P C	Chairman
2.	Dr. Renita Maria D'Souza	Member
3.	Dr. Raghavendra Rao, Associate Professor, PG Dept of Biotechnology, Alvas College, Moodbidri	Subject Expert
4.	Dr Shayama Prasad , Associate Professor, Dept of Biotechnology, NITTE, Karkala, Karnataka.	Subject Expert
5.	Dr Tharavathy N C, Professor, Dept. of Bioscience, Mangalore University, Karnataka	Vice Chancellor's nominee
6.	Mr Pavan Pai, Administrator	Representative from

	Shri Laxmi Narasimha Rice & Oil Mills company,Mangalore, Karnataka.	Industry
7.	Dr. Lyned Lasrado	Meritorious Alumnus
8.	Dr Shreelaitha Suvarna, Head, Dept. of PG Studies and Research in Biotechnology, St Aloysius College (Autonomous), Mangalore, Karnataka.	Special Invitees
9.	Miss Anindita Jana	Student Representative

A meeting of the Board of Study in Biotechnology (UG) was held on February 10, 2023, at 10 a.m. in the Conference Hall (Arrupe Block) at St. Aloysius College, Mangalore, as well as in hybrid mode on the Microsoft Team Platform, to discuss and approve the Vth semester Biotechnology syllabus of St. Aloysius College (Autonomous), Mangalore.

The chairman welcomed and introduced the members of the board of studies, and then the members were invited to discuss the syllabus for the fifth semester of biotechnology.

The BOS members had a brainstorming discussion and interaction among themselves regarding each course in the curriculum.

After discussion, fruitful suggestions were appropriately incorporated into the curriculum of the BSc Biotechnology NEP syllabus. The members approved the syllabus for both theory and practical courses with minor modifications. Based on the suggestions given by the members, BOS resolved to recommend the following to the Academic Council for further approval.

Members present:

Sl.No	Name	Role	Signature
1.	Mr. Sreejesh P C	Chairman	Present
2.	Dr. Renita Maria D'Souza	Member	Present
3.	Dr. T. S. Keshava Prasad Professor and Deputy Director, Yenepoya Deemed to be University, Mangaluru.	Subject Expert	Present
4.	Dr. Smitha Hegde Professor and Deputy Director, Nitte University, Mangaluru.	Subject Expert	Present
5.	Dr. Chandra. M Professor & Chairperson, Department of Biosciences, Mangalore University	Vice Chancellor's nominee	Absent
6.	Mr Rohan Shetty Senior Scientist II, Thermo Fisher Scientific, Bengaluru, Karnataka.	Representative from Industry	Absent
7.	Mrs. Bhagya Lakshmi	Meritorious Alumnus	Present
8.	Dr Ninaad Lasrado Postdoctoral Research Fellow and Virology Group Manager, Harvard Medical School, Boston, USA.	Special Invitee (International member)	Present- Online
9.	Dr Prveen Kumar M. K Manager, Quality Control, Molbio Diagnostics Private Limited, Goa.	Special Invitees (Representative form Industry)	Present- Online Online
10.	Dr Shreelaitha Suvarna, Head, Dept. of PG Studies and Research in Biotechnology, St Aloysius College, Mangalore.	Special Invitees	Present
11.	Nidhi Shenoy BSc BtCZ	Student Representative	Present

Preamble:

St. Aloysius College is the first college in Dakshina Kannada District to start an undergraduate Biotechnology programme in the year 2001. Bachelor / Hons. Program in biotechnology offers the synergism of multiplicity of scientific disciplines and technologies including cell biology, genetics, microbiology, biochemistry, molecular biology, Recombinant DNA technology, fermentation technology, enzyme technology, bioinformatics, and more recently a variety of omics. Our objective is to provide enhanced skills, knowledge, and research aptitude to carry out higher studies, entrepreneurship or research, and development and employment in the various health, research, and industrial areas.

The new Learning Outcome- Based Curriculum (LOBC) in Biotechnology will help the teachers of the discipline to visualize the curriculum more specifically in terms of the learning outcomes expected from the students at the end of the instructional process. The outcome based education (OBE) proposed by UGC helps to promote the link of students with the society/industry such that majority of the students engage in socially productive activities during their period of study in the institutions and at least half of the graduate students will secure access to employment/self-employment or engage themselves in pursuit of higher education. The model curriculum envisages to cater to the developmental trends in higher education, incorporating multi-disciplinary skills, professional and soft skills such as teamwork, communication skills, leadership skills, time management skills and inculcate human values, professional ethics, and the spirit of Innovation / entrepreneurship and critical thinking among students and promote avenues for display of these talents, linking general studies with professional courses. Besides imparting disciplinary knowledge to the learners, curriculum should aim to equip the students with competencies like problem solving, analytical reasoning and moral and ethical awareness. Introduction of internship and appropriate fieldwork/case studies are embedded in the curriculum for providing wider exposure to the students and enhancing their employability.

Learning outcomes specify what exactly the graduates are expected to know after completing a program of study. The expected learning outcomes are used as reference points to help formulate graduate attributes, qualification descriptors, program learning outcomes and course learning outcomes. Keeping the above objectives of higher education in mind the Learning Outcome-Based Curriculum Framework (LOCF) for the discipline of Biotechnology has been prepared. In keeping with the Govt of India's NEP-2020 vision of a holistic and multidisciplinary Under-Graduate education, has now been adopted in St Aloysius College (Autonomous), Mangalore

with minor changes and shall be effective from the academic year 2021-22.

Curriculum for B.Sc. (Basic/Hons.) Biotechnology

Program Name	B.Sc. Discipline	Total Credits for the Program	176
Core	Biotechnology	Starting year of implementation	2021-22

Salient features are as follows:

- Discipline Core (DSC) or Domain-specific Core Courses in Biotechnology as Major.
- Discipline Electives (DSE) or Elective Courses in the Core Subject or Discipline.
- Open Electives (OE) are Elective Courses offered to students from non-core Subjects across disciplines.
- Skill Enhancement Courses (SEC) that are domain-specific or generic.
- 1 hour of Lecture or 2 hours of practical per week in a semester is assigned one credit. Core discipline theory courses are of 3/4 credits, while practical are of 2 credits.
- Competencies need to be acquired by a candidate securing B.Sc. (Basic) or B.Sc. (Hons) degree in Biotechnology.

Program Outcomes:

By the end of the program the students will be able to:

PO 1. Understand concepts of Biotechnology and demonstrate interdisciplinary skills acquired in cell biology, genetics, biochemistry, microbiology, and molecular biology.

PO 2. Apply the knowledge and skills gained in the fields of plant biotechnology, animal biotechnology and microbial technology in pharma, food, agriculture, beverages, herbal and nutraceutical industries.

PO 3. Critically analyze environmental issues and apply the biotechnology knowledge gained for conserving the environment and resolving environmental problems.

PO 4. Demonstrate comprehensive innovations and skills in the fields of biomolecules, molecular biology, enzyme technology, bioprocess engineering and genetic

engineering of plants, microbes, and animals with respect to applications for human welfare.

- PO 5. Apply the knowledge and skills of immunology, bioinformatics, computational modelling of proteins, drug design and simulations to test models and aid in drug discovery.
- PO 6. Critically analyze, interpret data, and apply tools of bioinformatics and multi-omics in various sectors of biotechnology including health and food.
- PO 7. Demonstrate communication skills, scientific writing, data collection and interpretation abilities in all the fields of biotechnology.
- PO 8. Learn and practice professional skills in handling microbes, animals and plants and demonstrate the ability to identify ethical issues related to recombinant DNA technology, genetic engineering, animals handling, intellectual property rights, biosafety, and biohazards.
- PO 9. Explore the biotechnological practices and demonstrate innovative thinking in addressing the current day and future challenges with respect to food, health, and environment.
- PO 10. Demonstrate thorough knowledge and application of good laboratory and good manufacturing practices in biotech industries.
- PO 11. Apply the molecular biology principles and techniques in forensic and clinical biotechnology.
- PO 12. Demonstrate entrepreneurship abilities, innovative thinking, planning, and setting up of small-scale enterprises or CROs.

Programme Structure for BSc (Basic/Hons.) Biotechnology

Sem.	Discipline Core (DSC) (Credits) (L+T+P)	Discipline Elective (DSE) / Open Elective (OE) (Credits) (L+T+P)	Ability Enhancement Compulsory Courses (AECC), Languages (Credits) (L+T+P)		Skill Enhancement Courses (SEC)		Total Credits
					Skill based (Credits) (L+T+P)	Value based (Credits) (L+T+P)	
I	DSC-A1(4), A2(2) DSC-B1(4), B2(2)	OE-1 (3)	L1-1(3), L2-1(3) (4 hrs each)	Env. Studies (3) (3+0+0) OR	SEC-1: Digital Fluency (2) (1+0+2)	Health, Wellness & Yoga (2) (1+0+2)	25/26
II	DSC-A3(4), A4(2), DSC-B3(4), B4(2)	OE-2 (3)	L1-2(3), L2-2(3) (4 hrs each)	SEC-1: Digital Fluency (2) (1+0+2) OR	Env. Studies (3) (3+0+0)	Sports/NCC/NSS/R&R(S &G) / Cultural (2) (0+0+4)	26/25
Exit option with Undergraduate Certificate with completion of courses equivalent to a minimum of 48 credits, followed by an exit 10 credit skills enhancement course(s) for two months, including at-least 6-credit job-specific internship/apprenticeship to acquire job-ready competencies required to enter a job.							
III	DSC-A5(4), A6(2), DSC-B5(4), B6(2)	OE-3 (3)(3+0+0)	L1-3(3), L2-3(3) (4 hrs. each)		SEC-2: AI (2) (1+0+2)	Sports/NCC/NSS/R&R(S &G) / Cultural (2) (0+0+4)	25
IV	DSC-A7(4), A8(2), DSC-B7(4), B8(2)		L1-4(3), L2-4(3) (4 hrs. each)	India & Indian Constitution (3) (3+0+0)/ OR OE-3(3)	SEC-3: Financial Edu. & Inv. Aw. (2) (1+0+2)	Sports/NCC/NSS/R&R(S &G) / Cultural (2) (0+0+4)	25
Exit option with Undergraduate Diploma in Science Disciplines (with completion of courses equal to a minimum of 96 credits), followed by an exit 10 credit skills enhancement course(s) for two months, including at-least 6-credit job-specific internship/apprenticeship to acquire job-ready competencies required.							
V	DSC-A9(3), A10(2), A11(3), A12(2); DSC-B9(3), B10(2), DSC-B11(3), B12(2)	Vocational-1 (3)			SEC-4: Cyber Security (2)(1+0+2)/Internship (2)		25
VI	DSC-A13(3), A14(2), A15(3), A16(2); DSC-B13(3), B14(2), DSC-B15(3), B16(2)	Res.Methodology-1(3)			SEC-5: Relevant SEC (2) (1+0+2)/ Internship (2)		25
Exit option with the award of Bachelor of Science Degree, B.Sc. in Science Disciplines (with completion of courses equal to 132-140 credits).							

VII	DSC-A17(3), A18(2), A19(3), A20(2);	DSE-A1(3), A2(3) Res.Methodology-2(3)* Res. Proposal (4)*					23
VIII	DSE-A3(3), A4(3), A5(3), A6(3)	Res. Internship (4)* Research Project (6)*					22
Award of Bachelor of Science Degree with Honours, B.Sc. (Hons.) in Discipline A (with the completion of courses equal to a minimum of 176-180 credits)							

Scheme and Syllabus for B.Sc. (Basic / Hons.) Biotechnology

Semester	Course Category	Course Code	Title of Courses	Instruction hrs/week	Marks			Credits
					IA	SA	Total	
I	Discipline Specific Courses	G 511 DC1.1	Cell Biology and Genetics	4	40	60	100	4
		G 511 DC2.1P	Cell Biology and Genetics Practical	3	25	25	50	2
	Open Elective Courses*	G 511 OE1.1	Biotechnology for human welfare	3	40	60	100	3
II	Discipline Specific Courses	G 511 DC1.2	Microbiological methods and techniques	4	40	60	100	4
		G 511 DC2.2P	Microbiological methods and techniques Practical	3	25	25	50	2
	Open Elective Courses*	G 511 OE1.2	Applications of Biotechnology in agriculture	3	40	60	100	3
III	Discipline Specific Courses	G 511 DC1.3	Biomolecules	4	40	60	100	4
		G 511 DC2.3P	Biomolecules Practical	4	25	25	50	2
	Open Elective Courses*	G 511 OE1.3	Nutrition and Health	3	40	60	100	3
IV	Discipline Specific Courses	G 511 DC1.4	Molecular Biology	4	40	60	100	4
		G 511 DC2.4P	Molecular Biology Practical	3	25	25	50	2
	Open Elective Courses*	G 511 OE1.4	Intellectual Property Rights	3	40	60	100	3
V	Discipline Specific Courses	G 511 DC 1.5	Genetic Engineering	04	40	60	100	4
		G 511 DC 2.5P	Genetic Engineering Practical	04	25	25	50	2
		G 511 DC 3.5	Plant and Animal Biotechnology	04	40	60	100	4
		G511 DC 4.5P	Plant and Animal Biotechnology –Practical	04	25	25	50	2
VI	Discipline Specific Courses	G 511 DC 1.6	Immunology	04	40	60	100	4
		G 511 DC 2.6 P	Immunology Practical	04	25	25	50	2
		G 511 DC 3.6	Bioprocess and Environmental Biotechnology	04	40	60	100	4
		G 511DC 4.6P	Bioprocess and Environmental Biotechnology Practical	04	25	25	50	2

Pedagogical Note:

Pedagogy for student engagement is predominantly lectures. However, other pedagogies that enhance better student engagement may be adopted for each course. The list includes active/ experiential learning /course projects/ problem or project-based learning (PBL)/ case studies/ self- study like seminar, term paper or MOOC/ field visits / industrial visits / group activity / simulations / hackathons etc.

The general pedagogy to be followed for theory and practical are as follows: Lecturing, Tutorials, Group/Individual Discussions, Seminars, Assignments, Counseling, Remedial Coaching, Field/Institution/Industrial visits, Hands-on training, Case observations, Models/charts preparations, Problem solving mechanisms, Demonstrations, Project presentations, Experiential documentation, and Innovative methods, Active learning as per LSSSDC (NSDC) LFS/Q0509 (Lab Technician/Assistant-Life Sciences) guidelines, at skill training Level 3, Case studies.

Assessment: Every course needs to include assessment for higher order thinking skills (applying/ analyzing/evaluating/creating). These shall necessarily be reflected also in the Question Papers, such that questions of all levels of difficulty are framed. Alternate assessment methods that help formative assessment (i.e. assessment for learning) may also be adopted.

Syllabus for B.Sc. (Basic / Hons.)

SEMESTER – I

Discipline Specific Course.1

Course Title	CELL BIOLOGY AND GENETICS		
Course Code	G 511 DC1.1		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this course, students will be able to:

CO 1. Acquire a deep insight on the concepts of cell biology and describe the ultrastructure of cells, structure and function of organelles, cytosol and cytoskeleton.

CO 2. Illustrate the phases of cell cycle, cell division, reductional division in gametes, molecular mechanisms that regulate life and death of a cell including programmed cell death or apoptosis and differentiation in plants.

CO 3. Comprehend the organization and structure of chromosomes, banding patterns, karyotyping to detect genetic disorders and laws of inheritance, and gene interaction.

CO 4. Describe mutations and its types, genetic or hereditary disorders and prenatal screening of genetic disorders.

Unit 1. Cell as a basic unit of living systems and cellular organelles (14 hours)

Concept, Development and Scope of Biotechnology. Historical perspectives. Discovery of cell, the cell theory, Ultra structure of a prokaryotic and eukaryotic cell (Both plant and animal cells), Surface Architecture: Structural organization and functions of plasma membrane and cell wall of eukaryotes. Cellular Organelles: Structure and functions of cell organelles – Endoplasmic reticulum, Golgi complex, Mitochondria, Chloroplast, Ribosomes, Lysosomes, Peroxisomes, Nucleus (Nuclear envelope with nuclear pore complex, Nucleolus, Nucleoplasm and Chromatin). Vacuole, Cytosol and Cytoskeleton structures (Microtubules, Microfilaments, and Intermediate filaments).

Unit II. Chromosomes and cell division (14 hours)

General Introduction, Discovery, Morphology, and structural organization – Centromere, Secondary constriction, Telomere, Chromonema, Euchromatin and Heterochromatin, Chemical composition (molecular organization of chromosome and nucleosome model), Classification of chromosomes based on centromere position and

Karyotyping (methods). Giant Chromosomes: Salivary gland and Lamp brush chromosomes.

Cell Division: Cell cycle, phases of cell division, Mitosis and meiosis, regulation of cell cycle, cell cycle checkpoints, and enzymes involved in regulation, Significance of cell cycle, achromatic apparatus, synaptonemal complex. Cell Senescence and programmed cell death. Cell cycle disruption and Cancer.

Unit III. Laws of inheritance and gene interaction (14 hours)

Terminologies in genetics: alleles, gene, genome, Genotype, Phenotype, character, traits, homozygous and heterozygous.

Mendelian Laws of inheritance, monohybrid and dihybrid inheritance, test cross, back cross, law of segregation & independent assortment. Gene interactions: supplementary genes - Comb patterns in fowls, Complementary genes - Flower colour in sweet peas. Epistasis- Plumage colour in poultry. Chromosome theory of inheritance- Concept of allosomes and autosomes. Sex-linked inheritance (Haemophilia, Colour blindness), Multiple allelism: Blood groups in Human beings. OMIM database.

Unit IV Human genetics and medical genetics (14 hours)

Linkage and crossing over: Introduction, Coupling and repulsion hypothesis, Mechanism of crossing over and its importance.

Mutations: Types of mutation (somatic and germline mutation, Spontaneous: Point Mutations- Silent mutations, Missense mutations, Nonsense mutations, Frame shift mutation, transition and transversion; and induced, Mutagens: Physical, chemical and biological, Mutations in plants, animals, and microbes for economic benefit of man (one example each).

Human Genetics: A general account of structural and numerical aberrations, inherited disorders – Allosomal (Klinefelter syndrome and Turner's syndrome), Autosomal (Down's syndrome and Cri-Du-Chat Syndrome). Metabolic disorders due to inborn errors in metabolism (inborn errors associated with glucose metabolism Albinism, Alkaptonuria, Phenylketonuria, SCID). Prenatal screening for genetic disorder- Noninvasive methods - Maternal serum screening, Ultrasonography and Fetal echocardiography, Invasive methods-, Amniocentesis, Chorionic villus sampling. Genetic counselling.

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Acquire a deep insight on the concepts of cell biology and describe the ultrastructure of cells, structure and function of organelles, cytosol and cytoskeleton.	✓				✓							
CO 2. Illustrate the phases of cell cycle, cell division, reductional division in gametes, molecular mechanisms that regulate life and death of a cell including programmed cell death or apoptosis and differentiation in plants.	✓				✓				✓	✓		
CO 3. Comprehend the organization and structure of chromosomes, banding patterns, karyotyping to detect genetic disorders and laws of inheritance, and gene interaction.	✓								✓			
CO 4. Describe mutations and its types, genetic or hereditary disorders and prenatal screening of genetic disorders.	✓								✓			

References

1. Alberts, B., Hopkin, K., Johnson, A., Morgan, D., Lewis J., Raff M., Roberts, K., & Walter, P., (2019). Essential Cell Biology, International student edition 5th ed., WW Norton & Co.
2. Brooker, R.J., (2017). Genetic analysis and principle, 6th ed., Mc Graw Hill.
3. Cooper & Sinauer G.M., (2019). The Cell: A Molecular Approach, International 8th ed., Oxford University Press.
4. Hardin, J. & Bertoni, G P., (2018). Becker's World of The Cell, 9th ed., Pearson Education Ltd, USA.

5. Karp, G., Iwasa, J. & Marshall W., (2016). Cell and Molecular Biology: Concepts and Experiments, 8th ed., Wiley & sons. New York.
6. Lodish, H., Berk, A., Kaiser, C.A., Krieger, M., Bretscher, A., Ploegh, H., Amon, A. & Martin, K., (2016). Molecular Cell Biology, 8th ed., W.H. Freeman & Co., New York.
7. Gupta ML. and ML. Jangir. (2002) Cell Biology-Fundamentals and Applications. Argosies, Jodhpur, India.
8. Powar C.B.(2019). Cell Biology 3rd edition. Himalaya Publishing House, Mumbai.
9. Gardner, E.J., Simmons, M.J., Snustad, D.P. (2006). Principles of Genetics. VIII Edition John Wiley & Sons.
10. Gupta, P.K. (2019) Genetics, 5th Ed., Rastogi Publication, Meerut, India
11. Krebs, J.E., Goldstein, E.S. & Kilpartick, S.T., (2017). Lewin genes- XII, Jones and Bartlett Publishers.
12. Tamarin, R., (2017). Principles of Genetics, 7th ed., Mc-Graw – Hill Publication.
13. J. Brooker (2017). Genetics: Analysis and Principles., 6th ed., McGraw-Hill Education

SEMESTER – I

Discipline Specific Course.2

Course Title	CELL BIOLOGY AND GENETICS PRACTICAL		
Course Code	G 511 DC2.1P		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this Course, students will be able to:

- CO 1. Interpret the different stages of cell division and to calculate the mitotic index.
- CO.2. Measure the size of cells, count the number of cells using haemocytometer and perform the karyotyping analysis and solve various genetics problems.
- CO 3. Demonstrate the handling of *Drosophila melanogaster*, the model organism for genetic studies.
- CO 4. Describe the principles and procedures of genetic techniques in biological experiments

List of Practical

1. Handling and maintenance of simple and compound microscope
2. Use of Micrometry and calibration, measurement of onion epidermal cells and yeast.
3. Cell counting using haemocytometer.
4. Study of divisional stages in mitosis from onion root tips.
5. Determination of mitotic index in onion root tips.
6. Effect of osmotic pressure on RBC.
7. Study of divisional stages in meiosis in grasshopper testes/onion or Rheo flower buds.
8. Mounting of polytene chromosomes.
9. Buccal smear – Barr bodies.
10. Karyotype analysis – Human – Normal and Abnormal – Down and Turner's syndromes.
11. Mounting of the Sex Comb in *Drosophila melanogaster*.
12. Study of mutants in *Drosophila melanogaster*.
13. Separation of eye pigments of *Drosophila melanogaster*.
14. Genetic problems based on theory.

References

1. Vilas Parmar (2018). Practicals of Cell Biology & Genetics. LAP Lambert Academic Publishing.
2. Debarati D. (2017). Essential Practical Handbook of Cell Biology & Genetics, Biometry & Microbiology: A Laboratory Manual. Academic Publishers.
3. Amit Gupta and Bipin Kumar Sati (2019). Practical laboratory manual- Cell Biology. Lambert Academic Publishing.
4. Rina M. and Rama S. (2018). Laboratory Manual of Cell Biology. Prestige Publishers.

SEMESTER – I

OPEN ELECTIVE COURSE

Course Title	BIOTECHNOLOGY FOR HUMAN WELFARE		
Course Code	G 511 OE1.1		
Number of credits	3	Contact Hours	42

Course Outcomes:

After successful completion of this Course, students will be able to:

CO 1. Apply the biotechnological concepts in the industry.

CO 2. Implement the biotechnological techniques in environmental management.

CO 3. Describe application of biotechnology to forensic science.

CO 4. Comprehend contributions of biotechnology to biomedical fields, such as diagnostics, genomics, and therapeutics.

Unit 1. (14 hours)

Environment: Application of biotechnology in environmental aspects: Degradation organic pollutants – chlorinated and non-chlorinated compounds; degradation of hydrocarbons and agricultural wastes, PHB –production and its futuristic applications.

Unit II. (14 hours)

Industry: Application of biotechnology in industry: Industrial production of alcoholic beverages (wine), antibiotics (Penicillin), enzymes (lipase). Applications in food, detergent and pharmaceutical industry.

Unit III. Laws of inheritance and gene interaction (14 hours)

Forensic science: Application of biotechnology in forensic science: Solving crimes of murder and rape; solving claims of paternity and theft by using DNA finger printing techniques.

Health: Application of biotechnology in health: Genetically engineered insulin, recombinant vaccines, gene therapy, molecular diagnostics using ELISA and PCR. Monoclonal antibodies and their use in cancer. Human genome project.

References

1. Bhasin M.K. and Nath, S. (2002). Role of Forensic Science in the New Millennium, University of Delhi, Delhi
2. Crueger W. and Crueger A. (2000). Biotechnology: A textbook of Industrial Microbiology. 2nd Ed., Panima Publishing Co. New Delhi.

3. Eckert W.G. (1997) Introduction to Forensic Sciences, 2nd Ed., CRC Press, Boca Raton
4. James S.H. and Nordby, J.J. (2005). Forensic Science: An Introduction to Scientific and Investigative Techniques, 2nd Edition, CRC Press, Boca Raton
5. Joerdening H.-J. and Winter J. (2005). Environmental Biotechnology – Concepts and Applications,
6. Mohapatra, P.K. (2006) Textbook of Environmental Biotechnology, I.K. International Publishing House Pvt. Ltd., New Delhi
7. Nanda B.B. and Tiwari R.K. (2001). Forensic Science in India: A Vision for the Twenty First Century, Select Publishers, New Delhi
8. Patel A.H. (1996). Industrial Microbiology. 1st edition, Macmillan India Limited.
9. Stanbury P.F, Whitaker A and Hall S.J. (2006). Principles of Fermentation Technology. 2nd Ed., Elsevier Science Ltd.

SEMESTER – II

Discipline Specific Course.1

Course Title	MICROBIOLOGICAL METHODS AND TECHNIQUES		
Course Code	G 511 DC1.2		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this course, students will be able to:

CO 1. Employ the principles of microscopy to study microorganisms.

CO 2. Apply various analytical techniques and methods of sterilization in microbiological work.

CO 3. Delineate the formulation of media, culture methods and staining techniques for isolation, characterization of microbes.

CO 4. Apply the knowledge of antimicrobial agents in anti- microbial assays.

Unit 1. Introduction to microbes and methods to study (14 hours)

Classification of major groups of microorganisms- Bacteria, Fungi, Algae and viruses. binomial nomenclature of microbes, Phylogenetic classification; 16S rDNA sequencing. Microscopy: Principles of Microscopy- Magnification, resolving power, numerical aperture, working principle and applications and limitations of Compound microscope, Dark field microscope, Phase contrast microscope, Fluorescence Microscope, confocal microscope, Electron Microscopes- TEM and SEM. Analytical techniques: Working principles and applications: Centrifuge, Ultracentrifuge, UV-Vis Spectrophotometer, Chromatography- Paper and TLC.

Unit II. Sterilization techniques (14 hours)

Definition of terms-sterilization, disinfectant, antiseptic, sanitizer, germicide, microbicidal agents, microbiostatic agent and antimicrobial agent. Physical methods of sterilization: Principle, construction, and applications of moist heat sterilization by using autoclave, Pasteurization and Fractional sterilization-Tyndallization. Dry heat sterilization- hot air oven. Incineration. Filter sterilization- membrane filter and HEPA. Radiation- Ionizing radiation- γ rays and non-ionizing radiation- UV rays. Chemical methods: Alcohol, aldehydes, phenols, halogen, metallic salts, Quaternary ammonium compounds and sterilizing gases as antimicrobial agents.

Unit III. Culturing and staining methods**(14 hours)**

Culture Media: Nutritional types of bacteria, Components of media, Culture media types (natural and synthetic media, differential, enrichment and selective media).

Pure culture methods: Serial dilution and plating methods (pour, spread, streak).

Cultivation and maintenance of aerobic and anaerobic bacteria. Preservation/stocking of pure cultures: Agar slant cultures, agar slant culture covered with oil (Paraffin method), very low temperature (glycerol), Freeze Drying (lyophilization). Culture Collection Centres. Stains and staining techniques: Principles of staining, Types of stains-simple stains, structural stains and differential stains.

Unit IV. Antibacterial agents and Antibiotic resistance**(14 hours)**

Antibiotics and mode of action with one example each: Inhibitor of nucleic acid synthesis; Inhibitor of cell wall synthesis; Inhibitor of cell membrane function; Inhibitor of protein synthesis; Inhibitor of metabolism. Antibiotic resistance: MDR, XDR, MRSA, NDM-1. Antibiotic sensitivity testing methods: Kirby-Bauer method, Agar well diffusion techniques, and E-test, MIC.

Antifungal agents: Mechanism of action of Amphotericin B, Griseofulvin. Antiviral agents: Mechanism of action of Amantadine, Acyclovir, Azidothymidine.

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Employ the principles of microscopy to study microorganisms.	✓	✓										
CO 2. Apply various analytical techniques and methods of sterilization in microbiological work.		✓		✓								
CO 3. Delineate the formulation of media, culture methods and staining techniques for isolation, characterization of microbes.		✓		✓								
CO 4. Apply the knowledge of antimicrobial agents in anti- microbial assays	✓			✓					✓			

References

1. Black, J. G., & Black, L. J. (2017). Microbiology: Principles and Explorations, 10th ed., United States of America: John Wiley & sons, Inc.
2. Cann, A. J. (2016). Principles of Molecular Virology, 6th ed., London: Academic Press.
3. Dimmock, N. J., Easton, A. J., & Leppard, K. N. (2016). Introduction to Modern Virology, 7th ed., United Kingdom: Wiley-Blackwell.
4. Flint, J., Racaniello, V. R., Rall, G. F., & Skalka, A. M. (2015). Principles of Virology, 4th ed., Washington DC: ASM Press.
5. Madigan, M. T., Bender, K. S., Buckley, D. H., Sattley, W. M., & Stahl, D. A. (2019). Brock Biology of Microorganisms, 15th ed., Harlow, United Kingdom: Pearson.
6. Pommerville, J. C. (2011). Alcamo's Fundamentals of Microbiology, 9th ed., Sudbury, Massachusetts: Jones and Bartlett Publishers.
7. Talaro, K. P. (2009). Foundations in Microbiology: Basic Principles, 7th ed., New York: McGraw-Hill.
8. Tortora, G. J., Funke, B. R., & Case, C. L. (2015). Microbiology: An Introduction, 12th ed., United States of America: Pearson Education Inc.
9. Willey, J. M., Sherwood, L. M., & Woolverton, C. J. (2016). Prescott, Harley, and Klein's microbiology, 10th ed., Americas, New York: McGraw-Hill.
10. Pelczar Jr MJ, Chan ECS, and Krieg NR. (2004). Microbiology, 5th Ed., Tata McGraw Hill.
11. Atlas RM. (1997). Principles of Microbiology. 2nd edition. Wm C Brown Publishers.
12. Dubey R. C. and Maheshwari D. K. (2010). A Textbook of Microbiology. S Chand & Company
13. Ananthanarayan R, Jayaram Paniker CK and Reba Kanungo (2020). Textbook of Microbiology. 11th Ed. Universities Press (India) Pvt. Ltd.

SEMESTER – II

Discipline Specific Course.2

Course Title	MICROBIOLOGICAL METHODS AND TECHNIQUES PRACTICAL		
Course Code	G 511 DC2.2P		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this Course, students will be able to:

CO 1. Handle and use instruments used in microbiology and biotechnology laboratories.

CO 2. Experiment with various methods of sterilization in microbiological work

CO 3. Prepare different types of media, perform culture methods and staining techniques for isolation, characterization of microbes

CO 4. Handle and use antimicrobial agents and perform anti-microbial assays

List of Practical

1. Study the principle and applications of important instruments (biological safety cabinets, autoclave, incubator, hot air oven, compound microscope, pH meter) used in the microbiology laboratory.
2. Preparation of culture media for bacteria, fungi and their cultivation.
3. Isolation of bacteria and fungi from soil, water and air.
4. Enumeration techniques – direct microscopic, serial dilution and standard plate count technique (Spread plate, pour plate) and study of colony characters of isolated microbes.
5. Purification of bacterial cultures using streak plate technique.
6. Culture preservation techniques – slant and stab culture.
7. Study of colony characteristics bacteria
8. Staining techniques:-simple staining, Negative Staining, Gram's staining, Spore Staining - Schaeffer Fulton Method
9. Study of Bacterial motility by hanging drop technique.
10. Biochemical Tests for Bacterial Identification - IMViC test, catalase test, oxidase test
11. Antibiotic sensitivity test by disc diffusion method.
12. Fungi – Lactophenol / cotton blue staining.

13. Study of Rhizopus, Penicillium, Aspergillus using temporary mounts.

References

1. Cappucino J and Sherman N. (2010). Microbiology: A Laboratory Manual, 9th Ed., Pearson Education Limited.
2. Saha, Rumpa Das and Shukla (2021). Microbiology Practical Manual, 2nd Ed. CBS.
3. Mukesh Kumar (2018). Practical Manual for Undergraduates Microbiology, Jain Brothers.
4. Maheshwari D.K. and Dubey R.C. (2010). Practical Microbiology, S Chand & Company

SEMESTER – II

OPEN ELECTIVE COURSE

Course Title	APPLICATIONS OF BIOTECHNOLOGY IN AGRICULTURE		
Course Code	G 511 OE1.2		
Number of credits	3	Contact Hours	42

Course Outcomes:

After successful completion of this Course, students will be able to:

CO 1. Employ the biotechnological approaches in agriculture.

CO 2. Apply biotechnological methods in plant tissue culture.

CO 3. Comprehend the pros and cons of GM crops and their plant products.

Unit 1. (14 hours)

Agricultural Biotechnology: Concepts and scope of biotechnology in Agriculture. Plant tissue culture, micro propagation, entrepreneurship in commercial plant tissue culture.

Banana tissue culture – primary and secondary commercial setups, small scale bio-enterprises: Mushroom cultivation

Unit II. (14 hours)

Transgenic plants: The GM crop debate – safety, ethics, perception, and acceptance of GM crops GM crops case study: Bt cotton, Bt brinjal. Plants as bio factories for molecular

pharming; edible vaccines, plantibodies, nutraceuticals.

Unit III. Laws of inheritance and gene interaction (14 hours)

BT based pesticides: Baculovirus pesticides (NPV), Mycopesticides (Metarrhizium), Post-harvest Protection: Antisense RNA technology for extending shelf life of fruits and shelf life of flowers. Genetic engineering for quality improvement: Golden rice, Seed storage proteins (LEA), Flavours– capsaicin, vanillin

References

1. Chrispeels M.J. and Sadava D.E. (1994) Plants, Genes and Crop Biotechnology, 2nd Ed., Jones and Bartlett Publishers, Boston.
2. Gamborg O.L. and Philips G.C. (1998) Plant cell, tissue and organ culture, 2nd Ed., Narosa Publishing House. New Delhi.
3. Gistou, P. and Klu, H. (2004). Handbook of Plant Biotechnology (Vol. I & II). John Publication.
4. Hammond J., McGarvy P. and Yusibov.V. (2000). Plant Biotechnology, Springer Publ.
5. Heldt. H.-W. (1997). Plant Biochemistry and Molecular Biology. Oxford and IBH Publishing Co. Pvt. Ltd. Delhi.
6. Kyte, L., Kleyn, J., Scoggins, H., and Bridgen M. (2003) Plants from test tubes. An introduction to micropropagation, 4th Ed., Timber Press, Portland.
7. Murray D.R. (1996) Advanced methods in plant breeding and biotechnology. Panima Publishing Corporation.
8. Nickoloff, J.A. (1995). Methods in molecular biology, Plant cell electroporation and electrofusion protocols-Humana Press Incorp, USA.
9. Sawahel, W.A. (1997). Plant genetic transformation technology. Daya Publishing House, Delhi.

SEMESTER – III

Discipline Specific Course.1

Course Title	BIOMOLECULES		
Course Code	G 511 DC1.3		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this course, students will be able to:

CO 1. Cognise the properties of carbohydrates, proteins, lipids, cholesterol, DNA, RNA, glycoproteins and glycolipids and their importance in biological systems.

CO 2. Apprehend the importance of high energy compounds, electron transport chain, synthesis of ATP under aerobic and anaerobic conditions.

CO 3. Interpret the metabolic pathways such as Glycolysis, Kreb's Cycle, ETC, pentose phosphate pathway, etc. occurring inside living cells.

CO 4. Apply comprehensive innovations and skills of biomolecules to biotechnology field

Unit 1. Glycobiology and its metabolism (14 hours)

Carbohydrates: Classification of carbohydrates: Monosaccharides: structure of aldoses and ketoses, ring structure of sugars, conformations of sugars, mutarotation, anomers, epimers and enantiomers, structure of biologically important sugar derivatives.

Disaccharides: Glycosidic bond, reducing and nonreducing disaccharides, structure and functions of sucrose, lactose and maltose.

Polysaccharides: homo and heteropolysaccharides, structural and storage polysaccharides. structure and functions of starch, glycogen, and chitin. Glycosylation of other biomolecules: glycoproteins and glycolipids.

Metabolism of carbohydrates: Reactions, energetics, and regulation: Glycolysis, Fate of pyruvate under aerobic and anaerobic conditions, citric acid cycle, Pentose phosphate pathway, Gluconeogenesis. Mitochondrial electron transport chain, oxidative Phosphorylation. Energy balance of cellular oxidation of glucose.

Unit II. Lipids and its metabolism (14 hours)

Lipids: Structure and functions, Classification of lipids: simple and compound lipids. General structure and biological functions of: Phospholipids, sphingolipids, glycolipids, cerebrosides, gangliosides, Prostaglandins, and Cholesterol. Fatty acids: Classification:

short chain, medium chain, and long chain; saturated and unsaturated; essential fatty acids. Properties of fatty acids: acid number, Saponification number and iodine number.

Metabolism: Beta oxidation of fatty acids. Biosynthesis of cholesterol. Structure and properties of lipoproteins- HDL, LDL, VLDL.

Unit III. Amino acids and its metabolism (14 hours)

Structure and properties of amino acids. Classification of amino acids. Concept of – Zwitterion, isoelectric point, pK values. Essential and nonessential amino acids. Peptide bond and peptide, Forces stabilizing protein structure and shape, classification of proteins based on structure and function, Structural organization of proteins: primary, secondary (α , β), tertiary and quaternary. Fibrous and globular proteins. Denaturation and renaturation of proteins. General aspects of amino acid metabolism: Transamination, deamination, decarboxylation and urea cycle. Disorders of amino acids metabolism, phenylketonuria, alkaptonuria. Introduction to Proteomics.

Unit IV. Nucleic acid and its metabolism (14 hours)

Nucleic acids: structure of purines and pyrimidines, ribose and deoxy ribose, nucleoside and nucleotides, Chargaff's rule, Physical & chemical properties of nucleic acids: denaturation, renaturation, melting temperature, hyperchromicity, cot curve. RNA: types of RNA (t-RNA, r-RNA, m-RNA & micro-RNAs). Introduction to nucleic acid sequencing.

Nucleotide metabolism: Overview of *de novo* and salvage pathway of nucleotide synthesis, Inhibitors of nucleotide metabolism. Disorders of purine and pyrimidine metabolism: Lesch-Nyhan syndrome, Gout, SCID, adenosine deaminase deficiency.

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Cognise the properties of carbohydrates, proteins, lipids, cholesterol, DNA, RNA, glycoproteins and glycolipids and their importance in biological systems.	✓			✓			✓					
CO 2. Apprehend the importance of	✓			✓			✓					

high energy compounds, electron transport chain, synthesis of ATP under aerobic and anaerobic conditions.

CO 3. Interpret the metabolic pathways such as Glycolysis, Krebs's Cycle, ETC, pentose phosphate pathway, etc. occurring inside living cells. ✓ ✓ ✓

CO 4. Apply comprehensive innovations and skills of biomolecules to biotechnology field. ✓ ✓ ✓ ✓

References

1. Agarwal, G. R., & Agarwal, O. P. (2007). Textbook of Biochemistry. Krishna Prakashan Media.
2. Campbell, M. K., Farrell, S. O., & McDougal, O. M. (2016). Biochemistry. Cengage Learning.
3. Champe, P. C., Harvey, R. A., & Ferrier, D. R. (2005). Biochemistry. Lippincott Williams & Wilkins.
4. Cox, M. M., Nelson, D. L., Lehninger, A. L. (2008). Lehninger principles of biochemistry. United Kingdom: W. H. Freeman.
5. Hames, D., & Hooper, N. (2006). *Instant notes biochemistry*. Taylor & Francis.
6. Jain, J.L, Sunjay, J. & Nithin, J.(2012). Fundamentals of Biochemistry. 6th ed. S. Chand & Company.
7. Lehninger, A. L. (2012). Principles of Biochemistry 6th ed.. New York, NY: Worth.
8. Satyanarayana, U. (2017). Biochemistry. Elsevier Health Sciences.
9. Rodwell, V. W., Bender, D. A., Botham, K. M., Kennelly, P. J., & Weil, P. A. (2018). Harper's illustrated biochemistry. New York, NY, USA:: McGraw-Hill Education.
10. Voet, D., Voet, J. G., & Pratt, C. W. (2016). Fundamentals of biochemistry: life at the molecular level. John Wiley & Sons.

SEMESTER – III

Discipline Specific Course.2

Course Title	BIOMOLECULES PRACTICAL		
Course Code	G 511 DC2.3P		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this Course, students will be able to:

CO 1. Determine presence of biomolecules like carbohydrates, proteins, lipids, etc. in known and unknown samples.

CO 2. Determine the extent of adulteration in samples containing biomolecules.

CO 3. Construct the standard curve, analyze the data and interpret the results.

CO 4. Apply knowledge of biochemistry and metabolism in various cellular functions, and the application of research involved in various biochemical processes.

List of Practical

1. Preparation of buffers.
2. Verification of Beer's law- determination of absorbance maxima of proteins
3. Separation of carbohydrates by paper chromatography.
4. Separation of Amino acids by Thin layered chromatography.
5. Qualitative tests for Carbohydrates.
6. Qualitative tests for lipids .
7. Qualitative tests for amino acids and proteins
8. Estimation of reducing sugar by DNS method.
9. Estimation of Proteins by Biuret method.
10. Estimation of Proteins by Lowry's method.

References

1. Dhiman, P., Rajendiran, S., Dhiman, P., Rajendiran, S. (2019). Biochemistry Practical Manual - E-Book. India: Elsevier Health Sciences.
2. Plummer, D. T., Plummer, D. T. (2001). Introduction to Practical Biochemistry. India: Tata McGraw Hill Publishing Company.
3. Vasudevan, D., Das, K. S., Vasudevan, D., Das, K. S. (2019). Practical Textbook of Biochemistry for Medical Students. India: Jaypee Brothers Medical Publishers Pvt. Limited.

4. Wilson, K., & Walker, J. (Eds.). (2010). Principles and techniques of biochemistry and molecular biology. Cambridge university press.

SEMESTER – III

OPEN ELECTIVE COURSE

Course Title	NUTRITION AND HEALTH		
Course Code	G511 OE1.3		
Number of credits	3	Contact Hours	42

Course Outcomes:

CO 1. Apply the concepts of food, nutrition, and diet in life and design a balanced diet.

CO 2. Apply the best practices of food intake and dietary requirements.

CO 3. Acquire knowledge about various sources of nutrients and good cooking practices.

Unit 1. Introduction (14 hours)

Concepts of nutrition and health. Definition of food, diet and nutrition, food groups. food pyramids. Functions of food. Balanced diet. Meal planning. Eat right concept. Functional foods, Prebiotics, Probiotics, and antioxidants

Unit II. Nutrients (14 hours)

Macro and Micronutrients - Sources, functions and deficiency. Carbohydrates, Proteins, Fats – Sources and calories. Minerals –Calcium, Iron, Iodine. Vitamins – Fat soluble vitamins –A, D, E & K. Water soluble vitamins – vitamin C Thiamine, Riboflavin, Niacin. Water–Functions and water balance. Fibre –Functions and sources. Recommended Dietary Allowance, Body Mass Index and Basal Metabolic Rate.

Unit III. Nutrition and Health (14 hours)

Methods of cooking affecting nutritional value. Advantages and disadvantages. Boiling, steaming, pressure cooking. Oil/Fat – Shallow frying, deep frying. Baking. Nutrition through lifecycle. Nutritional requirement, dietary guidelines: Adulthood, Pregnancy, Lactation, Infancy- Complementary feeding, Pre-school, Adolescence, geriatric. Nutrition related metabolic disorders- diabetes and cardiovascular disease.

References

1. Sri Lakshmi B, (2007), Dietetics. New Age International publishers. New Delhi
2. Sri Lakshmi B, (2002), Nutrition Science. New Age International publishers. New Delhi
3. Swaminathan M. (2002), Advanced textbook on food and Nutrition. Volume I. Bappco
4. Gopalan.C., RamaSastry B.V., and S.C.Balasubramanian (2009), Nutritive value of Indian Foods.NIN.ICMR.Hyderabad.
5. Mudambi S R and Rajagopal M V, (2008), Fundamentals of Foods, Nutrition & diet therapy by New Age International Publishers, New Delhi.

SEMESTER – IV

Discipline Specific Course.1

Course Title	MOLECULAR BIOLOGY		
Course Code	G 511 DC1.4		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this course, students will be able to:

CO 1. Acquire a deep insight on the concepts of central dogma in Molecular biology.

CO 2. Describe the fine structure of DNA and the mechanism of replication in prokaryotes and eukaryotes.

CO 3. Comprehend the causes of DNA damage and various mechanism of DNA repair.

CO 4. Illustrate the fundamental principles of gene expression and regulation in cells.

Unit 1. DNA structure and replication (14 hours)

DNA as genetic material: Experiments of Griffith, Avery and Hershey & Chase. Structure of DNA, Types of DNA (A, B & Z), Organelle DNA: cpDNA and mtDNA. Chromatin and the Nucleosome, Histones and Non-Histones. Central dogma in molecular biology. Replication of DNA in prokaryotes: Semiconservative nature of DNA replication, replicon, origin of replication, enzymes in DNA replication. Unique aspects of eukaryotic chromosome replication, Fidelity of replication.

Unit II. DNA damage, repair, and homologous recombination (14 hours)

DNA damage: causes (physical and chemical) and types of DNA damage, mechanism of DNA repair: Photoreactivation, base excision repair, nucleotide excision repair, mismatch repair, recombinational repair, SoS repair, nonhomologous end joining. Homologous recombination- Holliday model.

Unit III. Transcription and RNA processing (14 hours)

RNA structure and types of RNA, transcriptome, Transcription in prokaryotes: Prokaryotic RNA polymerase, role of sigma factor, promoter, Initiation, elongation, and termination of RNA synthesis. Polycistronic mRNA.

Transcription in eukaryotes: Eukaryotic RNA polymerases, transcription factors, promoters, enhancers, mechanism of transcription initiation, promoter clearance and elongation. RNA splicing and processing: processing of pre-mRNA: 5' cap formation, polyadenylation, splicing of mRNA: spliceosome. RNA editing and role of guide RNA.

Inhibitors of transcription: Rifampicin, α -amanitin.

Unit IV. Translation and regulation of gene expression (14 hours)

Genetic code and its characteristics. Translation in Prokaryotes and Eukaryotes: ribosome structure and assembly, charging of tRNA, aminoacyl tRNA synthetases, Mechanism of initiation, elongation and termination of polypeptides, Post translational modifications of proteins.

Regulation of gene expression in prokaryotes: Operon concept, Lac operon-fine structure, repressor, and the catabolite activator in regulation of lactose operon. Transcriptional control by attenuation in tryptophan operon. Gene regulation in eukaryotes: galactose operon in yeast.

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Acquire insight on the concepts of central dogma in Molecular biology.	✓			✓			✓	✓				
CO 2. Describe the fine structure of DNA and the mechanism of replication in prokaryotes and eukaryotes.	✓			✓			✓	✓			✓	
CO 3. Comprehend the causes of DNA damage and various mechanism of DNA repair.	✓			✓			✓				✓	
CO 4. Illustrate the fundamental principles of gene expression and regulation in cells.	✓			✓			✓	✓			✓	

References

1. Alberts B., Hopkin K., Johnson A., Morgan D., Lewis J., Raff M., Roberts K. and Walter P., (2019), Essential Cell Biology, International student edition 5th ed., WW Norton & Co.
2. Brown T. A., (2017), Genomes 4, 4th ed., Garland Science, Taylor and Francis group, New York.

3. Cooper & Sinauer G.M., (2019), The Cell: A Molecular Approach, International 8th ed., Oxford University Press
4. De Robertis, E.D.P. and De Robertis, E.M.F. (2017). Cell and Molecular Biology. VIII Edition. Lippincott Williams and Wilkins, Philadelphia.
5. Karp G., Iwasa J. & Marshall W., (2016), Cell and Molecular Biology: Concepts and Experiments, 8th ed., Wiley & sons. New York.
6. Lodish, H., Berk, A., Kaiser, C.A., Krieger, M., Bretscher, A., Ploegh, H., Amon, A. & Martin, K., (2016), Molecular Cell Biology, 8th ed., W.H. Freeman & Co., New York
7. P.S. Verma, D. V. A. (2009). Molecular Biology. S. Chand Limited., India
8. Tropp B.E., (2020), Molecular Biology: Genes to Proteins, 5th ed., Jones & Bartlett Learning.
9. Watson, J. D., Baker T.A., Bell, S. P., Gann, A., Levine, M., and Losick, R., (2014) Molecular Biology of the Gene (VII Edition.). Cold Spring Harbour Lab. Press, Pearson Pub.
10. Wilson and Walker (2018). Principles and Techniques of Biochemistry and Molecular Biology. United Kingdom: Cambridge University Press.

SEMESTER – IV

Discipline Specific Course.2

Course Title	MOLECULAR BIOLOGY PRACTICAL		
Course Code	G 511 DC2.4P		
Number of credits	4	Contact Hours	56

Course Outcomes:

After successful completion of this Course, students will be able to:

CO 1. Independently execute laboratory experiments using the standard methods and techniques in molecular biology, with the appropriate analysis and interpretation of results obtained.

CO 2. Independently use various instruments such as centrifuges, colorimeters, UV-transilluminator, Gel Doc, UV- Vis spectrophotometer in laboratory work.

List of Practical

1. Isolation of chromosomal DNA from yeast / bacterial cells.
2. Isolation of genomic DNA from animal tissues/ plant tissues.
3. Estimation of DNA by diphenylamine method.
4. Isolation of RNA from coconut endosperm/ yeast.
5. Estimation of RNA by orcinol method.
6. Isolation of Plasmid DNA by alkaline lysis method.
7. Agarose gel electrophoresis of genomic DNA & plasmid DNA
8. Separation of proteins and Molecular weight determination by SDS-PAGE.
9. Quantification and purity analysis of DNA by UV spectrophotometer.
10. SDS PAGE

References

1. Chaitanya, K. V. (2013). Cell and Molecular Biology: A Lab Manual. PHI Learning, India.
2. Fernandez, T. G., Pattison, S. (2015). Biochemistry Laboratory Manual For Undergraduates: An Inquiry-Based Approach. De Gruyter., Germany.
3. Gakhar, S. K., Miglani, M., Kumar, A. (2013). Molecular Biology: A Laboratory Manual. I.K. International Publishing House Pvt. Limited, India.
4. Lone, S. M., Rasool, R. S., Masoodi, K. Z. (2020). Advanced Methods in Molecular Biology and Biotechnology: A Practical Lab Manual. Elsevier Science., Netherlands.

5. Miller, H. B., Carson, S., Witherow, D. S., Carson, S. (2012). Molecular Biology Techniques: A Classroom Laboratory Manual.: Elsevier Science., Netherlands.
6. Ravi, I., Baunthiyal, M., Saxena, J. (2015). Laboratory Manual of Microbiology, Biochemistry and Molecular Biology. Scientific Publishers., India.
7. Thompson, D. (2011). Cell and Molecular Biology Lab Manual. CreateSpace Independent Publishing Platform.

SEMESTER – IV
OPEN ELECTIVE COURSE

Course Title	INTELLECTUAL PROPERTY RIGHTS		
Course Code	G511 OE1.4		
Number of credits	3	Contact Hours	42

Course Outcomes:

CO 1. Elucidate different types of intellectual property rights in general and protection of products derived from biotechnology research.

CO 2. Acquire knowledge about filing patents, process, and infringement.

CO 3. Evaluate multiple perspectives about trademarks, industrial designs, and copyright.

Unit 1. Introduction to Intellectual property rights (IPR) (14 hours)

Genesis and scope. Types of Intellectual property rights - Patent, Trademarks, Copyright, Design, Trade secret, Geographical indicators, Plant variety protection. National and International agencies – WIPO, World Trade Organization (WTO), Trade-Related Aspects of Intellectual Property Rights (TRIPS), General Agreement on Tariffs and Trade (GATT).

Unit II. Patenting, process, and infringement (14 hours)

Basics of patents - Types of patents; Patentable and Non-Patentable inventions, Process and Product patent. Indian Patent Act 1970; Recent amendments; Patent Cooperation Treaty (PCT) and implications. Process of patenting. Types of patent applications: Provisional and complete specifications; Concept of “prior art”, patent databases (USPTO, EPO, India). Financial assistance, schemes, and grants for patenting. Patent infringement- Case studies on patents (Basmati rice, Turmeric, Neem).

Unit III. Trademarks, Copy right, industrial Designs (14 hours)

Trademarks- types, Purpose and function of trademarks, trademark registration, Protection of trademark. Copy right- Fundamentals of copyright law, Originality of material, rights of reproduction, industrial Designs: Protection, Kind of protection provided by industrial design.

References

1. Ashok, K. M. & Mohd, I. A. (2008). Intellectual property rights. 1st Ed. Serials Publications New Delhi.
2. Acharya, N.K. (2014). Textbook of intellectual property rights. 7th Ed. Asia Law House, Hyderabad.
3. Fleming D.A., Hunt D. (2000) Biological safety Principles and practices 3rd Ed. ASM Press.

4. Ganguli, P. (2001). Intellectual property rights: Unleashing the knowledge economy. New Delhi: Tata McGraw-Hill Pub.
5. Manish Arora. (2007). Universal's Guide to Patents Law 4th Ed. Universal Law Publishing House.
6. Kalyan C. Kankanala. 2012. Fundamentals of Intellectual Property. Asia Law House
7. World trade organization - <http://www.wto.org>
8. World Intellectual Property organization – www.wipo.int Office of the controller general of Patents, Design & Trademarks - www.ipindia.nic.in.

SEMESTER – V

Discipline Specific Course.1

Course Title	GENETIC ENGINEERING		
Course Code	G511 DC1.5		
Number of credits	4	Contact Hours	60
Course Outcome:			

After successful completion of this course, students will be able to:

CO 1. Describe the principles, role of various DNA modifying enzymes used in genetic engineering.

CO 2. Implement a range of methodologies for genetic manipulation, encompassing diverse practices and genome editing techniques.

CO 3. Utilise various genetic engineering practices such as PCR, Hybridisation methods for diagnosis and various advanced development.

CO 4. Explore the multifaceted applications of genetic engineering in the biotechnology industry, employ computational tools in genetic manipulation, and critically examine the ethical implications associated with the manipulation of genetic material and its societal impact.

Unit I. Fundamentals of Genetic Engineering (15 hours)

Introduction to rDNA technology. Isolation and purification of DNA (genomic and plasmid) and plant and animal DNA. Methods for quantification and characterization of DNA samples.

Restriction-modification systems: Restriction enzymes-types, isoschizomers and neoschizomers. double digests, Restriction enzymes – function, classification (Based on recognition and restriction sequence: -type I, II and III; based on pattern of restriction: sticky and blunt end cutters, Restriction mapping. DNA modifying enzymes and its functions (DNA Polymerases, Klenow fragment, DNA Ligase, S1 Nuclease, Alkaline Phosphatase, Terminal Transferase, Polynucleotide kinases, Polynucleotide phosphorylase, RNase A, RNase H, DNase 1, DNase II, Exonuclease III, Reverse Transcriptase).

Essential features of cloning and expression vectors. Selectable markers and reporter genes. Plasmid vectors: pUC19, pBR322. M-13 vectors and Cosmid vectors. Artificial chromosome vectors (YAC, BAC). Animal vectors: SV40 vectors and Adeno-associated virus (AAV) vectors, baculoviruses as vectors. Plant vectors: Agrobacterium plasmid biology: organization of Ti plasmid and T-DNA transfer, cointegrate and binary vectors.

Unit II. Practices in Genetic Engineering (15 hours)

Cohesive and blunt end ligation; Linkers; Adaptors; Homopolymer tailing. Construction of libraries: genomic library; Isolation of mRNA and methods of cDNA synthesis and construction of cDNA library. Gene knockout techniques in bacterial and eukaryotic organisms.

Physical and chemical methods of gene transfer: Biolistic, Microinjection, electroporation, Calcium phosphate co precipitation, Liposome mediated transformation, PEG, DEAE dextran.

Identifying the right clones: Insertional inactivation of marker gene, visual screening- Blue-white selection, colony and plaque hybridization, Immunological techniques.

DNA sequencing- Sanger's, Next Generation Sequencing. RNA sequencing and its types. Genome editing technology: Principles and applications of genome editing techniques, CRISPER/Cas 9, site-directed mutagenesis, and other genome editing methods.

Unit III. Applications of Genetic Engineering (15 hours)

Analysis of gene expression: Southern blot, Northern blot and western blot. In situ hybridization: FISH & GISH. Recombinant Protein Expression and Purification, affinity tags in protein purification.

PCR: Basic principle and amplification of DNA, Types of PCR – multiplex, nested, reverse transcriptase, real time PCR.

DNA fingerprinting and its applications in forensics. Molecular diagnostic techniques and their role in disease diagnosis. Use of genetic engineering in the development of therapeutics and vaccines. Production of biopharmaceuticals using recombinant DNA technology. Genomics: methods and applications.

Unit IV. Advances in Genetic Engineering (15 hours)

Industrial applications of genetic engineering, such as enzyme production, biofuel production, and bioremediation. Introduction to synthetic biology and its integration with genetic engineering. Design and construction of artificial biological systems. Bioinformatics and Computational Tools: Introduction to bioinformatics and its role in genetic engineering. Use of computational tools for sequence analysis, gene prediction, and protein structure analysis.

Ethical and Regulatory Considerations - Discussion of ethical implications associated with genetic engineering. Introduction to regulatory guidelines and safety considerations for genetic engineering research and applications

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Describe the principles, role of various DNA modifying enzymes used in genetic engineering.				✓				✓				
CO 2. Implement a range of methodologies for genetic manipulation, encompassing diverse practices and genome editing techniques.	✓			✓	✓			✓		✓	✓	
CO 3. Utilise various genetic engineering practices such as PCR, Hybridisation methods for diagnosis and various advanced development.	✓			✓				✓			✓	
CO 4. Explore the multifaceted applications of genetic engineering in the biotechnology industry, employ computational tools in genetic manipulation, and critically examine the ethical implications associated with the manipulation of genetic material and its societal impact.				✓		✓	✓	✓	✓		✓	✓

References

14. Watson J.D. (2007). Recombinant DNA technology: genes and genomes 3rd edition. W.H. Freeman and company
15. Brown T. A., (2019). Gene Cloning and DNA Analysis: An Introduction, 7th ed. Blackwell Science (UK). ISBN: 978-1119072560
16. Gene Cloning and DNA Analysis: An Introduction (2019) 7th ed., Brown, TA, Wiley Blackwell,
17. Brown T. A., (2017). Genomes 4 , 4th ed., Garland Science, Taylor and Francis group, NewYork.

-
18. Huss, Mikael, Korpelainen, Eija, Somervuo, Panu, Tuimala, Jarno, Wong, Garry (2017). RNA-seq data analysis a practical approach. CRC Press, Taylor & Francis Group.
 19. Micklos D.A., & Hilgert U. (2013). Genome science: A practical and conceptual introduction, Cold Spring Harbour Press.
 20. Pawan K.D. and Vijai S.,(2020) Genome Engineering Via CRISPR-Cas9 System, Elsevier Science & Technology.
 21. Rastogi and Pathak. (2013). Genetic engineering, Oxford University Press.
 22. Roberto Biassoni, Alessandro Raso (2020). Quantitative Real-Time PCR: Methods and Protocols. 2ed Humana press.
 23. Lewin B. (2010). Genes X ed., Oxford Univ Press.
 24. Twyman & Primrose. (2009). Principles of gene manipulation, Butterworth Heinemann.
 25. Watson J.D., Baker T.A. & Bell A.P. (2013). Molecular Biology of the Gene, 7th ed., Cold Spring Harbor Laboratory Press.
-

SEMESTER – V
Discipline Specific Course.2

Course Title	GENETIC ENGINEERING PRACTICAL		
Course Code	G 511DC1.5 P	Number of hours /weeks- 4	
Number of credits	2	Contact Hours	60
Course Outcome:			

After successful completion of this course, students will be able to:

CO 1. Utilize essential genetic engineering tools and techniques effectively for conducting practical experiments.

CO.2 Design experiments for isolation, purification and amplification of DNA for genetic engineering work.

CO 3. Analyse and interpret genetic data using bioinformatics tools for understanding of gene function and evolutionary relationships.

List of Practical:

1. Isolation of nucleic acids from different sources.
2. Quantification of DNA by UV-Vis spectroscopy
3. Restriction enzyme digestion of DNA and calculation of molecular weight of the digested DNA
4. Isolation of Plasmids-electrophoretic identification of linear, circular and supercoiled DNA
5. DNA ligation
6. DNA amplification by PCR.
7. Preparation of competent cells in *E. coli*.
8. Calcium chloride mediated transformation of *E. coli* & selection of transformants
9. Demonstration experiments: Southern Blotting.
10. Restriction mapping
11. Bioinformatics for Genetic Engineering

Introduction to bioinformatics databases and tools

Sequence analysis (e.g., BLAST, multiple sequence alignment) Prediction of protein structure and function.

12. Gel Electrophoresis and DNA Analysis

Agarose gel electrophoresis for DNA fragment separation and analysis DNA size

determination using molecular weight markers. DNA band visualization techniques (e.g., ethidium bromide staining, DNA intercalating dyes).

References

1. Abdelmigid, H. (2013). Genetic Engineering: Laboratory Manual. (n.p.): CreateSpace Independent Publishing Platform.
2. Masoodi, K. Z., Lone, S. M., Rasool, R. S. (2020). Advanced Methods in Molecular Biology and Biotechnology: A Practical Lab Manual. Netherlands: Elsevier Science.
3. Noble. (2023). A Complete Lab Manual for Biotechnology. (n.p.): Notion Press.
4. Kurnaz, I. A. (2015). Techniques in Genetic Engineering. United Kingdom: CRC Press.

SEMESTER – V
Discipline Specific Course. 3

Course Title	PLANT AND ANIMAL BIOTECHNOLOGY		
Course Code	G 511 DC2.5		
Number of credits	4	Contact Hours	60
Course Outcome:			

After successful completion of this course, students will be able to:

CO 1. Analyze the fundamental concept of cellular totipotency and adeptly apply this understanding to proficiently execute micropropagation techniques, including bud culture, meristem culture, and organogenesis.

CO 2. Evaluate the applications of transgenic plants and analyze their safety and ethical concerns.

CO 3. Apply the basic techniques of mammalian cell culture, including parameters for measuring cell growth, viability, and cytotoxicity.

CO 4. Analyze the concepts of gene therapy and cloning, evaluate applications of transgenic animals, and consider their ethical implications.

Unit I. Plant Tissue culture

(15 hours)

Introduction to plant tissue culture, history, definition, Concept of cellular totipotency, cytodifferentiation - xylogenesis, Commercial PTC laboratory organization; Brief overview of nutrient requirements, culture media (MS media), role of growth regulators, and factors influencing the growth. Basic steps involved in plant tissue culture selection of mother plant, explant, surface sterilisation of explant. Callus. Differentiation: Organogenesis and somatic embryogenesis, synthetic seeds. Micropropagation, bud culture and meristem culture. Haploid Production: anther culture, pollen culture, ovary culture, ovule culture. Embryo culture, Meristem culture and production of virus free plants.

Secondary metabolites, *in vitro* secondary metabolite production, Cell suspension cultures, growth vs secondary metabolite production, bioreactors and scaling up of secondary metabolite production, limitations, and applications.

Unit II. Transgenic Plants and biosafety

(15 hours)

Overview of transgenic plants and their significance in agriculture. Applications of Transgenic Plants - Improved crop traits through genetic engineering: pest resistance (Bt gene- case study of Bt -cotton), herbicide tolerance, disease resistance: bacteria and fungi (pathogenesis related proteins, anti-microbial proteins, phytoalexins), and virus

(coat protein mediated cross protection,), and abiotic stress tolerance. Case study of golden rice and Flavr Savr tomato.

Biosafety assessment of transgenic plants: potential risks and benefits. International/national regulatory frameworks for releasing and commercializing genetically modified organisms (GMOs). Ethical and socio-economic impacts of transgenic crops. Intellectual property rights and access to transgenic technologies.

Unit III. Animal Cell culture methods (15 hours)

History and laboratory organisation (Equipment and materials for animal cell culture), Media components and types. Cell types and culture characters. Pluripotency, Multipotency, Differentiation, Trans differentiation, Reprogramming.

Biology and characterization of cultured cells- cell adhesion, proliferation, differentiation, morphology of cells, and identification. The basic technique of mammalian cell culture *in vitro*, Measuring parameters of growth in cultured cells, cell viability, and cytotoxicity. Large-scale culture of cell lines- monolayer, suspension, and immobilized cultures.

Organ and histotypic culture: Technique, advantages, limitations, applications. Stem cells: types (embryonic, adult, induced pluripotent), isolation, identification, expansion, differentiation and uses, stem cell engineering.

Unit IV. Gene therapy and transgenic animals. (15 hours)

Gene therapy: Somatic therapy and germline therapy with examples – SCID, CF. Tissue engineering and applications (e.g. artificial skin).

Reproductive cloning (nuclear transplantation- Cloning of Dolly) and therapeutic cloning (xenotransplantation).

Biopharming: concept, mammary glands of farm animals as bioreactors for production of regulatory proteins [α - anti trypsin (AAT), human tissue plasminogen activator], Silkworms as bioreactors for production of heterologous proteins. Transgenic animals and applications (e.g. transgenic cattle, sheep and fish). CPCSEA guidelines.

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Analyze the fundamental concept of cellular totipotency and	✓	✓								✓		✓

adeptly apply this understanding to proficiently execute micropropagation techniques, including bud culture, meristem culture, and organogenesis.												
CO 2. Evaluate the applications of transgenic plants and analyze their safety and ethical concerns.		✓		✓	✓	✓		✓	✓			
CO 3. Apply the basic techniques of mammalian cell culture, including parameters for measuring cell growth, viability, and cytotoxicity.		✓				✓				✓		✓
CO 4. Analyze the concepts of gene therapy and cloning, evaluate applications of transgenic animals like biopharming and xenotransplantation, and consider their ethical implications.		✓	✓			✓		✓				✓

References

1. Chawla, H. S. (2000). Introduction to Plant Biotechnology. Enfield, NH: Science.
2. Razdan, M. K. (2003). Introduction to Plant Tissue Culture. Enfield, NH: Science.
3. Slater, A., Scott, N. W., & Fowler, M. R. (2008). Plant Biotechnology: an Introduction to Genetic Engineering. Oxford University Press.
4. Buchanan, B. B., Gruissem, W., & Jones, R. L. (2015). Biochemistry & Molecular Biology of Plants. John Wiley & Sons.
5. Glick, B.R., and Pasternak, J.J. (2018). Molecular Biotechnology: Principles and Applications of Recombinant DNA. 5th edition. Washington, DC: ASM Press.
6. Satyanarayana, B.N. & Varghese, D.B.(2007). Plant tissue culture practices and new experimental protocols, IK International Publications.
7. Razdan, M. (2016). Introduction to Plant Tissue Culture 3rd edition. Oxford University Press.

-
8. Neal Stewart Jr. (2016). Plant Biotechnology and Genetics: Principles, Techniques and Applications. Wiley and Sons, New York.
 9. Taiz, L., and Zeiger, E. (2014). Plant Physiology. 5th edition. Sunderland, MA: Sinauer Associates.
 10. Freshney, I. (2016). Culture of Animal Cells: A Manual of Basic Technique and Specialized Applications (8th ed.). Wiley-Blackwell.
 11. Pörtner, R. (Ed.). (2007). Animal Cell Biotechnology: Methods and Protocols. Humana Press.
 12. Singh, B., & Gautam, S.K. (2013). Textbook of Animal Biotechnology. The Energy and Resources Institute (TERI).
 13. Gupta, P.K. (2018). Animal Biotechnology. Rastogi Publications.
 14. Mather, J.P., & Barnes, D. (Eds.). (Year N/A). Animal Cell Culture Methods. In Methods in Cell Biology, Vol. 57. Academic Press.
 15. Singh, B.D. (2006). Biotechnology: Expanding Horizons (3rd ed.). Kalyani Publishers.
 16. Srivastava A.K. Animal Biotechnology. (2018). Oxford & IBH Publishing Co Pvt.Ltd.
-

SEMESTER – V

Discipline Specific Course.4

Course Title	PLANT AND ANIMAL BIOTECHNOLOGY PRACTICAL		
Course Code	G 511 DC2.5P		
Number of credits	2	Contact Hours	60

Course Outcome:

After successful completion of this course, students will be able to:

CO 1. Demonstrate skills in plant and animal tissue culture techniques.

CO 2. Apply knowledge of cell and tissue culture techniques and PCR for both research and commercial purposes.

CO 3. Plan and conduct experiments with the use of specific methods applied to *in vitro* culture of animal cell .

List of Practical:

1. Preparation of MS media for plant tissue culture.
2. Seed culture
3. Callus culture form *Daucus carota*
4. Preparation of synthetic seeds- sodium alginate-CaCl₂ method.
5. Explant preparation – Leaf, bud, rhizome, and meristem
6. Micropropagation in Bougainvillea plant
7. Anther culture
8. Acclimatization and hardening techniques
9. Preparation of cell culture media: Preparation of basic cell culture media, such as Dulbecco's Modified Eagle Medium (DMEM), supplemented with fetal bovine serum (FBS), antibiotics, and other required additives.
10. Filter sterilization: Practice filter sterilization for sensitive media ingredients
11. Primary explant culture using liver cells / kidney / spleen cells.
12. Disaggregation of liver tissue by Warm Trypsin method.
13. Estimation of cell viability for the trypsinized liver cells by dye exclusion method

14. Assessing cell viability using the MTT assay.

References:

1. Santosh Nagar, Madhavi Adhav (2009). Practical Biotechnology and Plant Tissue Culture. S. Chand Limited.
2. Chawla. H.S. (2004). Plant Biotechnology: Laboratory Manual for Plant Biotechnology. Oxford & IBH Publishing Company Pvt. Limited.
3. Reinert J., Yeoman M.M. (2012). Plant Cell and Tissue Culture, A Laboratory Manual. Springer Berlin Heidelberg
4. Park, S. (2021). Plant Tissue Culture: Techniques and Experiments. Netherlands: Elsevier Science.
5. John Davis. (2011). Animal Cell Culture: Essential Methods. United Kingdom: Wiley.
6. Capes-Davis, A., Freshney, R. I. (2021). Freshney's Culture of Animal Cells: A Manual of Basic Technique and Specialized Applications. United Kingdom: Wiley.
7. Freshney, R. I. (2015). Culture of Animal Cells: A Manual of Basic Technique and Specialized Applications. Germany: Wiley.

SEMESTER – VI
Discipline Specific Course. 1

Course Title	IMMUNOLOGY
Course Code	G 511 DC1.6
Number of credits	4
Contact Hours	60

After successful completion of this course, students will be able to:

CO 1. Explain the distinction between innate and acquired immunity and recognize the roles of B and T-lymphocytes in both humoral and cell-mediated immune responses.

CO 2. Understand major histocompatibility complexes, antigen processing pathways, complement pathways, cytokines, and types of hypersensitivity reactions.

CO 3. Apply knowledge of immunodiffusion reactions, agglutination reactions, ELISA, RIA, immunocytochemistry, and fluorescent techniques in diagnosis.

CO 4. Apply the domain-specific knowledge and skills acquired in immunology for innovative therapies and Immuno-technologies.

Unit I. Cells and Organs of the Immune System (15 hours)

Introduction to the Immune System: History of Immunology, Types of Immunity: first and second line of defense, innate and acquired/adaptive immunity, specificity, diversity.

Cells of the immune system: Antigen-presenting cells (APCs), Role of B and T-lymphocytes in Humoral immunity and cell-mediated immunity, primary and secondary immune response, Immunization, memory. Organs of the Immune system: Thymus, bone marrow, spleen, Lymph Node, peripheral lymphoid organs.

Unit II. Molecules of the Immune System (15 hours)

Antigens and haptens: Properties (foreignness, molecular size, heterogeneity). Adjuvants. Antigenicity and Immunogenicity. Affinity and Avidity. B and T cell epitopes, superantigens Immunoglobulins: Classification, structure, and function. Antibody diversity (Neutralising and Non neutralising antibodies) , Monoclonal and polyclonal antibodies.

Major histocompatibility complexes: Classification, structure, and function. Antigen processing pathways – Cytosolic and Endocytic, Complement Pathways, Cytokines: Classification and function, Hypersensitivity: Reactions – Types I, II, and III. Delayed Type Hypersensitive Response.

Unit III. Immunotechniques and vaccines (15 hours)

Structure and properties of antigens- iso- and allo-antigens, antigen specificity, Cross-

reactivity, Precipitation, Immunodiffusion reactions: Radial immunodiffusion, Ouchterlony double diffusion, Immuno electrophoresis. Agglutination: Agglutination reactions. Use of antibody conjugates in bioassays: ELISA, RIA, Immunocytochemistry. Vaccines: Conventional (killed or attenuated vaccines), peptide vaccines, subunit, DNA vaccines, mRNA vaccines. Toxoids, antisera, edible vaccines, plantibodies, and Cancer vaccines.

Unit IV. Immunology of diseases

(15 hours)

Transplantation immunology: Phases in graft rejection and immuno-suppressors. Autoimmune Disorders: Systemic and Organ-specific Autoimmune disorders with examples. Immunodeficiencies: Primary and secondary immunodeficiencies; acquired immunodeficiency syndrome.

Cancer and the immune system – immune surveillance, immunological escape, cancer antigens, cancer immunotherapy.

Microbial diseases in humans: Mode of infection, symptoms, epidemiology and control measures of diseases caused by Viruses (Hepatitis-B), Bacteria (Typhoid), Fungi (Aspergillosis), Protozoa (Malaria).

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Explain the distinction between innate and acquired immunity and recognize the roles of B and T-lymphocytes in both humoral and cell-mediated immune responses.					✓		✓					
CO 2. Understand major histocompatibility complexes, antigen processing pathways, complement pathways, cytokines, and types of hypersensitivity reactions.					✓		✓					
CO 3. Apply knowledge of immunodiffusion reactions, agglutination reactions, ELISA, RIA,		✓			✓		✓		✓		✓	✓

immunocytochemistry, and fluorescent techniques in diagnosis.												
CO 4. Apply the domain-specific knowledge and skills acquired in immunology for innovative therapies and Immuno-technologies.		✓			✓	✓	✓		✓			

References

1. Abbas A, Lichtman, A. H. & Pillai, S., (2017), Cellular and Molecular Immunology, 9th ed., Elsevier.
2. Coico, R. & Sunshine, G., (2015), Immunology: A Short Course, 7th ed., Wiley-Blackwell.
3. Delves, P.J., Martin, S.J., Burton, D. R. & Roitt, I. M., (2017), Roitt's Essential Immunology, 13th ed., Wiley- Blackwell.
4. Kindt, T. J., Goldsby, R. A., Osborne, B. A. & Kuby, J., (2007), Kuby Immunology, 7th ed. 2007. W.H. Freeman.
5. Lippincott W.E., (2014), Fundamental Immunology, 7th ed. Paul, Williams and Wilkins
6. Male, D., Brostoff, J., Roth, D. B. & Roitt, I. V., (2012), Elsevier Immunology, 8th ed.
7. Murphy, K. & Weaver, C., (2016), Janeway's Immunobiology, 9th ed., Garland Science.
8. Owen J., Jenni Punt J. & Stranford S., (2018), Kuby Immunology, 8th ed., Jones P. W. H. Freeman.
9. Playfair, J. H. L. & Chain, B. M., (2012), Immunology at a Glance, 10th ed., Wiley-Blackwell.
10. Tizard, I. R., (2000), Immunology: an Introduction, 4th ed., Cengage Learning India.
11. Ananthanarayan, R., Paniker, C. J. (2006). Ananthanarayan and Paniker's Textbook of Microbiology. India: Orient Longman Private Limited.

SEMESTER – VI
Discipline Specific Course. 2

Course Title	Immunology Practical		
Course Code	G 511 DC2.6P		
Number of credits	2	Contact Hours	60

Course Outcome:

After successful completion of this course, students will be able to:

CO 1. Execute immune cell differential counting and whole count of white blood cells accurately.

CO 2. Successfully perform immunoassay techniques such as radial immunodiffusion and Ouchterlony double diffusion, dot ELISA, serum immune-electrophoresis, and Western blotting.

CO 4. Demonstrate proficiency in analyzing and interpreting experimental results.

List of Practical:

1. Hemagglutination of ABO Blood groups
2. Determination of Rh factor
3. Whole Count of WBC using Hemocytometer
4. Cells of the Immune System- Differential counting by Giemsa/Leishman stain.
5. Radial immunodiffusion
6. Ouchterlony double diffusion
7. Testing for Typhoid antigens by Widal test
8. Dot ELISA
9. Serum Immunoelectrophoresis.
10. Western Blotting
11. Cell culture techniques-Chinese Hamster Ovary (CHO) Cells
12. Estimation of viability and enumeration of the CHO cells.

References:

1. Sam-Yellowe, T. Y. (2021). Immunology: Overview and Laboratory Manual. Germany: Springer International Publishing.
2. Speshock, J. (2019). Immunology Lab Manual. (n.p.): Kendall Hunt Publishing Company.

3. Arora, B., Arora, D. R. (2020). Practical Microbiology. India: CBS Publishers & Distributors.
4. Maheshwari D.K. (2002). Practical Microbiology. India: S. Chand Limited.

SEMESTER – VI
Discipline Specific Course. 3

Course Title **Bioprocess and Environmental Biotechnology**

Course Code **G 511 DC3.6**

Number of credits **4** **Contact Hours** **56**

Course Outcome:

After successful completion of this course, students will be able to:

CO 1. Describe the principles of upstream processing and calculate growth rates and biomass yields within diverse culture modes.

CO 2. Compare and contrast various bioreactor designs and describe the principles and applications of various downstream processing techniques.

CO 3. Propose biotechnological solutions to address specific environmental pollution challenges.

CO 4. Design a bioremediation strategy for a specific contaminant and propose methods for effective wastewater treatment and solid waste management.

Unit I. Introduction to bioprocess technology (14 hours)

Basic principle components of fermentation technology. Strain improvement of industrially important microorganisms (membrane permeability, metabolic engineering, auxotrophic mutants, analogue resistant mutants, Use of recombination systems). Types of microbial culture and its growth kinetics– Batch, Fed-batch, and Continuous culture. Principles of upstream processing – Design of fermentation media (Carbon and nitrogen sources, precursors and metabolic regulators, Anti-foams), Inocula development, and sterilization -thermal death kinetics. sterilization of medium (batch and continuous).

Unit II. Bioreactors and downstream processing (14 hours)

Bioreactors- Design of a bioreactor, Significance of Impeller, Baffles, Sparger; Specialized bioreactors- design and their functions: airlift bioreactor, tubular bioreactors, membrane bioreactors, tower bioreactors, fluidized bed reactor, packed bed reactors.

Downstream processing- cell disruption, precipitation methods, solid-liquid separation, liquid-liquid extraction, filtration (Rotary vacuum filters, membrane filtration- micro filtration, reverse osmosis and ultrafiltration), centrifugation (principle, Basket and tubular bowl), chromatography (GPC, IEC, Affinity), drying devices (Lyophilization and spray dry technology), crystallization, biosensors-

construction and applications, Microbial production of ethanol, Penicillin, and Single Cell Proteins.

Unit III. Fundamentals of Environmental Biotechnology- (14 hours)

Air, water and soil microbiology

Introduction to Environmental Biotechnology- soil microorganisms-Interaction among microorganisms in Soil: Positive and Negative interactions: Neutralism, Commensalisms, Synergism (proto-cooperation), Mutualism (symbiotic), Competition, Amensalism, Parasitism and Predation.

Microbial composition of air, Sampling Techniques for trapping of indoor and air borne microbes in brief: agar plate, Gravity slide. Anderson, Burkard.

Air borne diseases in brief (Tuberculosis, Corona, SARS, MERS) and allergens (Hay fever, Rhinitis).

Analysis of Water –sampling, qualitative (Presumptive, Confirmed and completed coliform test) and quantitative -Membrane filter technique. Standards of water quality for drinking and industry.

Water borne Diseases: Water borne pathogens and diseases- Bacterial (Cholera, Shigella) and Protozoal (Amoebiasis, Giardiasis).

Role of Biotechnology in Environmental Conservation. Biotechnological Methods in Pollution detection and Abatement. Reduction of CO₂ emission using biotechnological approaches. Application of cell immobilization techniques in pollution abatement.

Unit IV. Biofertilizers, Biopesticides, Bioremediation and (14 hours)

Waste Management

Introduction to biofertilizers, Production of biofertilizers and utilization of organisms- for Biological Nitrogen fixation. Ex: *Rhizobia*, cyanobacteria, *Azotobacter*, *Azospirillum*, Phosphate solubilizing organisms, mycorrhiza- ectomycorrhiza and endomycorrhiza, sea weeds for soil enrichment

Introduction to biopesticides, properties, organisms- bacteria (*Bacillus thuringiensis*, *Bacillus papillae*, *Bacillus sphaericus*), Fungi (*Beauveria bassiana*) virus (*Baculovirus*), protozoans and plant products as biopesticides. Limitations of biopesticides.

Importance of bioremediation in environmental cleanup. Types (Phyto and microbial bioremediation) and microorganisms used in bioremediation. *In-situ* and *ex-situ* Bioremediation Methods. Microbial degradation of Xenobiotics

(pesticides, hydrocarbons and detergents) and bio- mining.

Wastewater Management- Primary (screening and sedimentation), secondary (trickling filters, activated sludge process, oxidation ponds, rotating biological contactors, fluidized bed reactor), tertiary, advanced treatment of wastewater. Anaerobic Digestion and Biogas Production. Solid Waste Management- Vermi composting, organic waste treatment using black soldier fly.

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs)	Program Outcomes (POs)											
	1	2	3	4	5	6	7	8	9	10	11	12
CO 1. Describe the principles of upstream processing and calculate growth rates and biomass yields within diverse culture modes.	✓	✓		✓			✓	✓		✓		
CO 2. Compare and contrast various bioreactor designs and describe the principles and applications of various downstream processing techniques.				✓		✓	✓		✓	✓		✓
CO 3. Propose biotechnological solutions to address specific environmental pollution challenges.			✓				✓		✓			
CO 4. Design a bioremediation strategy for a specific contaminant and propose methods for effective wastewater treatment and solid waste management.			✓				✓	✓	✓			✓

References

1. Stanburry, P.F. & Whitaker, (2005). 2nd ed., Principles of Fermentation Technology, Pergamon Press, Butterworth Heinemann-Elsevier.
2. Stanbury, P. F., Whitaker, A., Hall, S. J. (2016). Principles of Fermentation Technology. Netherlands: Elsevier Science.

-
3. Arindam, K., Vinay, S. (2018). Principles and Applications of Fermentation Technology. United States: Wiley.
 4. Das, D., Pandit, S. (2021). Industrial Biotechnology. United States: CRC Press.
 5. Okafor, N., Okeke, B. C. (2017). Modern Industrial Microbiology and Biotechnology. United Kingdom: CRC Press.
 6. Kalaichelvan, P.T., Arul, P. (2019). Bioprocess Technology. (n.p.): MJP Publisher.
 7. Shuler M.L. & Kargi F. (2002). Bioprocess Engineering Basic Concepts 2nd ed., Prentice Hall of India pvt. Ltd.
 8. Patel AH. (1996). Industrial Microbiology. 1st edition, Macmillan India Limited.
 9. Peavy, H. S., Rowe, D. R., & Tchobanoglous, G. (2014). Environmental engineering. McGraw-Hill Education.
 10. Banerjee, S., & Santhosh, C. (2019). Environmental biotechnology: Concepts and applications. CRC Press.
 11. Mishra, A. K. (2016). Environmental biotechnology: Basic concepts and applications. CRC Press.
 12. Mohapatra, P. K. (2013). Textbook of Environmental Biotechnology. India: I.K. International Publishing House Pvt. Limited.
-

SEMESTER – VI
Discipline Specific Course. 3

Course Title **Bioprocess and Environmental Biotechnology Practical**

Course Code **G 511 DC4.6P**

Number of credits **2** **Contact Hours** **56**

Course Outcome:

After successful completion of this course, students will be able to:

CO 1. Utilize crowding plate method to isolate industrially important microorganisms from natural resources.

CO 2 Apply various techniques to estimate key water quality parameters including sulphates, phosphates, chlorides, biological oxygen demand and microbial analysis of water.

CO 3. Produce amylase, ethanol, lactic acid, and wine through fermentation processes.

CO 4. Gain hands-on experience in essential techniques such as ammonium sulphate precipitation, dialysis, and total solids (TS) determination.

List of Practical:

1. Isolation of industrially important microorganisms from natural resources.
(Crowded plate and screening for amylase production)
2. Submerged fermentation and analysis of amylase
3. Ammonium sulphate precipitation
4. Dialysis
5. Study of fermentor- Demonstration.
6. Production of wine
7. Estimation of the percentage of alcohol, total acidity & volatile acidity in wine.
8. Production and analysis of ethanol.
9. Production and analysis of lactic acid.
10. Mushroom cultivation-demo.
11. Determination of TS and TDS
12. Microbial analysis of water-MPN, Confirmed and Completed test.
13. Estimation of Sulphates
14. Estimation of Phosphates

15. Determination of DO and BOD in water.

16. Estimation of chlorides.

References:

1. Patra, J. K., Das, G., Das, S. K., Thatoi, H. (2020). A Practical Guide to Environmental Biotechnology. Germany: Springer Nature Singapore.
2. Kulandaivel, S., Janarthanan, S. (2012). Practical Manual on Fermentation Technology. India: I.K. International Publishing House Pvt. Limited.
3. Chellapandi, P. (2007). Laboratory Manual in Industrial Biotechnology. India: Pointer Publishers.
4. Mamta, B., Indu, R., Jyoti S. (2019). Comprehensive Laboratory Manual of Life Sciences. (n.p.): Scientific Publishers.

Internship/ Mini project for graduate Programme

Course title	Internship Discipline specific
Course code	
No of contact hours	90
No credits	2
Method of evaluation	Presentations/Report submission/Both

Project Assessment			
Formative Assessment		Summative Assessment	Total Marks
Assessment Occasion/ type	Weightage in Marks	Practical Exams	
Data maintenance	10	Presentation/Report/Both 25	50
Assessment	10		
Attendance	05		
Total	25	25	

- Internship shall be Discipline Specific of 90 hours (2 credits) with duration 4-6 weeks.
- Internship shall be domain specific industry internship or academic research project.
- Internship may be full-time/part-time (full-time during semester holidays and part-time in the academic session)
- Internship mentor/supervisor shall avail work allotment during 6th semester for a maximum of 20 hours.
- The student should submit the final internship report (90 hours of Internship) to the

mentor for completion of the internship.

**Question Paper Pattern for End Semester Theory Examination
(Same scheme to be followed for all Semesters)**

Program: B.Sc. Biotechnology

Name of Course:

Course code:

Time: 2.5 Hours

Max. Marks: 60

Note: Draw neat, labelled diagrams wherever necessary

Part -A

- I Answer any **FIVE** of the following. **(5x 2=10)**
(Short answer questions any **FIVE** to be answered out of **eight**)

Part-B

- II Answer any **SIX** of the following (any **SIX** to be answered out of **(6 x 5 = 30)**
Eight)

Part-C

- III Answer any **TWO** of the following (any **TWO** to be answered **(02x10=20)**
out of **Four**)

Part A: Short answer questions shall be based on basic, conceptual, understanding etc.

Part B: Critical notes / Descriptive questions shall be based on deeper understanding, analytical, problem-solving skills etc.

Part C: Essay type questions shall be on critical thinking, higher order thinking skills etc.

Continuous Internal Assessment for theory (40 Marks)

Components:	IA Marks
Continuous Internal Assessment (Two internal tests 10 x 2)	20 marks
Assignment	05 marks
Attendance/Regularity	05 marks
Surprise test/ Open book exam/ Unit wise test (Objective/MCQ)/Seminar.	05 marks
Group Project work/ MOOC course/ Poster or Paper presentation	05 marks

Continuous Internal Assessment for Practical (25 Marks)

Assessment Occasion / Components	IA Marks
Continuous Internal Assessment of all practical experiments	05 marks
Test	10 marks
Record	05 marks
Attendance	05 marks

Question paper Pattern for practical examination. (Same scheme to be followed for all Semesters)

End semester Practical exam.

Program: B.Sc. Biotechnology

Name of Course:

Course code:

Time: 3 Hrs		Total marks: 25
I	Major experiment	12 marks
II	Minor experiment	8 marks
III	Spotters A, B, C (3 x 1 marks)	3 marks
IV	Viva	2 Marks
